

Aqueous extract effects of some common weed species against certain plant pathogenic fungi

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تأثيرات المستخلصات المائية لبعض الأعشاب الدخيلة على الفطريات المسببة لأمراض نباتية.

تم فحص التأثيرات المثبطة لأربعة وستون نوعا عشبيا شائعا تنتمي لتسعة وعشرون عائلة نباتية ضد فطر البنيسليوم ديجيتاتوم وفطر سكلوروتينيا سكلوروشوروم وفطر فيرتيسيليوم داليا وذلك تحت ظروف المختبر. وقد أظهرت النتائج أن المستخلصات العشبية مختلفة في تأثيراتها المضادة للفطريات وفي مثابة تلك التأثيرات، كما أن التأثيرات المضادة للفطريات تختلف من نوع عشبي إلى آخر ومن عائلة نباتية لأخرى. ومن بين جميع الأنواع العشبية التي تم فحصها فقد لوحظ أن مستخلصات عشب الرومراوم والحلاوي والحوذان هي الأكثر تنبيطا لفطر السكلوروتينيا سكلوروشوروم في حين أن مستخلصات الخلبوب والليزيق والحويرنة والحوذان كانت الأشد تنبيطا لنمو وتطور فطر الفيرتيسيليوم داليا. ولقد لوحظ أن المستخلصات المائية لعشب الحوذان كانت الأكثر تسما للأنواع الفطرية الثلاث حيث منعت بشكل كامل نموها وإنتاج الأبواغ الفطرية طيلة فترة الحضانة.

الكلمات المفتاحية : السمية للفطريات - مستخلصات - بنيسليوم ديجيتاتوم - سكلوروتينيا سكلوروشوروم - فيرتيسيليوم داليا - الأعشاب.

Effet des extraits aqueux de certaines adventices sur les champignons phytopathogènes

Les effets fongicides de 64 adventices fréquentes appartenant à 29 familles botaniques contre *Penicillium digitatum* Sacc., *Sclerotinia sclerotiorum* (Lib) De Bary et *Verticillium dahliae* Kleb ont été étudiés dans les conditions du laboratoire. Les extraits aqueux de plusieurs espèces de mauvaises herbes sont toxiques vis-à-vis de la croissance et/ou la sporulation des champignons étudiés. Cependant, les extraits aqueux de certaines espèces ont un effet stimulateur. L'activité fongicide et la persistance des extraits varient en fonction de l'espèce et de la famille botanique. *Chenopodium murale*, *Crepis aspera* et *Ranunculus asiaticus* ont des extraits plus toxiques sur *P. digitatum*. Cependant, les extraits d'*Enodium cruciatum*, *Euphorbia helioscopia*, *R. asiaticus* sont plus nocifs vis-à-vis de *S. sclerotiorum* alors que ceux d'*E. helioscopia*, *Galium tricornutum*, *Sisymbrium irio*, *R. asiaticus* sont plus nocifs vis-à-vis de *V. dahliae*. Les extraits de *R. asiaticus* ont été plus toxiques vis-à-vis des trois champignons étudiés (croissance et sporulation complètement inhibées durant toutes les périodes d'incubation).

Mots clés : Effet fongicide - Extraits - *Penicillium digitatum* - *Sclerotinia sclerotiorum* - *Verticillium dahliae* - Adventices

Aqueous extract effects of some common weed species against certain plant pathogenic fungi

Antifungal effects of 64 common weed species belonging to 29 plant families against *Penicillium digitatum* Sacc., *Sclerotinia sclerotiorum* (Lib.) De Bary, and *Verticillium dahliae* Kleb were investigated under laboratory conditions. Aqueous extracts of many weed species were toxic to growth and/or sporulation of one more of the tested fungi, while certain weed extracts showed stimulatory effects. Extracts were varied in their antifungal activities and in the persistence of their effects. Antifungal effects differ from species to species and from family to another. Among all species tested, the most toxic extracts were those of *Chenopodium murale*, *Crepis aspera* and *Ranunculus asiaticus* to *P. digitatum*. *Erodium cruciatum*, *Euphorbia helioscopia* and *R. asiaticus* to *S. sclerotiorum* and *E. helioscopia*, *Galium tricornutum*, *Sisymbrium irio* and *R. asiaticus* to *V. dahliae*. The extract from *R. asiaticus* was the most toxic to the three fungal species, and completely prevented their growth and sporulation at all periods of incubation.

Keys words : Fungi toxicity - Extracts - *Penicillium digitatum* - *Sclerotinia sclerotiorum* - *Verticillium dahliae* - Weeds

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INTRODUCTION

The important role that plants may play as a source of natural chemicals and their importance in controlling different agricultural pests is well documented (Frange, 1984 ; Rice, 1986 ; Narwal, 1994). Many workers have reported antifungal activity of plant extracts, volatile materials or oils against different plant pathogenic fungi (Frange, 1984 ; Lapis & Dumancas, 1978 ; Guesin & Reveillera, 1984 ; Akhtar *et al.*, 1986 ; Al Abed *et al.*, 1993) and recent studies on the subject emphasised the importance of natural chemicals as an alternative to synthetic pesticides in any future strategy for pest control (Beye, 1978 ; Macias *et al.*, 1994 ; Singh, 1994).

Studies on the antifungal activities of plant species found in Jordan are in their infancy. Work on the possible antifungal effects of extracts of some of the most widely spread weed species is important in the search for less hazardous, cheaper and more selective chemicals.

MATERIALS AND METHODS

The effect of weed species on the growth and development of *Penicillium digitatum* Sacc. (causing green mold disease) from orange fruits, *Sclerotinia sclerotiorum* (Lib.) De Bary from eggplant shoots (causing cottony stem rot) and *Verticillium dahliae* Kleb from watermelon roots (causing wilt disease) was investigated under laboratory conditions. The shoots of 64 weed species belonging to 29 plant families were collected from different locations in the country and their aqueous extracts were tested for mycotoxic effects.

• Preparation of weed extracts

The shoots of different weed species were cut at various growth stages (Table 1). Samples (300 g) of the fresh shoots of each specie were first washed in running tap water and distilled water, then the plant material was placed in distilled water (1 L) and the mixture homogenized, using a Waring Blender, for five minutes. The mixture was then allowed to stand for half an hour. The supernatant was filtered through Whatman N° 1 filter paper followed by a membrane filter (0.2 µm to remove any bacterial or fungal contamination).

Fungi species selected for this study were *Penicillium digitatum* Sacc. from orange fruits,

Sclerotinia sclerotiorum (Lib.) De Bary from eggplant shoots and *Verticillium dahliae* Kleb from watermelon roots.

Four discs (1 cm diameter) of each fungi specie, taken from 8 day old cultures, were placed in sterile Petri-dishes (11 cm diameter) containing 20 ml of sterile potato dextrose agar (PDA) medium and 3 ml extract of each weed species. The control treatment was 3 ml of sterile distilled water without any extract added.

The Petri-dishes were incubated for 16 days at 20°C (except for *V. dahliae* at 18°C) in the dark and then a visual estimation on the growth and spore formation of the tested fungi species, using 1-10 scale (with the lower score being most effective) was taken at 4, 8 and 16 days after incubation.

Treatments were arranged in a complete randomized block design with 4 replicates, all data were subjected to analysis of variance (ANOVA) and treatment means were compared using least significant differences (L.S.D. ; $P < 0.05$).

RESULTS

Aqueous extract effects of different weed species on growth of the three fungi are shown in Table 1.

Many weed species showed antifungal activity against one or more fungi species. Extracts were different in the strength of their phytotoxic effects.

At 4 days of incubation, 52 weed species showed antifungal activity against *P. digitatum* and their extracts did significantly reduce its colony growth compared with the control. At 8 days of incubation, extracts of 20 species significantly reduced fungal growth while at 16 days, there were 18 species that showed inhibitory effect on this fungus. The most effective extracts were those of *C. murale*, *D. erocoides*, *P. rhoeas*, *R. asiaticus*, *S. irio* and *V. cruciatum* and at all periods of incubation.

Extracts of *C. iberica*, *C. murale*, *E. helioscopia*, *R. asiaticus*, *S. nigrum*, *S. palaestinum* and *V. cruciatum* strongly inhibited growth of *S. sclerotiorum* at all incubation periods. Colony growth reduction was more than 70% of the control. *E. cruciatum*, *Lamium sp.* and *S. arvensis* showed lower inhibitory effects.

Similar differences in the antifungal effects of extracts were obtained with *V. dahliae*.

Table 1. Visual estimation of the effect of shoot extracts of different weed species on the growth of *P. digitatum*, *S. Sclerotiorum* and *V. dahliae* at three dates of incubation. Nomenclature is that of Flora Pauciflora, Zohary (1960) (continued)

Family and Scientific name/weed	Growth stage	<i>P. digitatum</i>			<i>Sclerotium</i>			<i>V. dahliae</i>		
		Days of incubation			Days of incubation			Days of incubation		
		4	6	10	4	6	10	4	6	10
Labiace										
<i>Matricaria inodora</i> C. Presl	Vegetative	17	30	30	60	63	33	60	13	17
<i>Lamium</i> sp.	Flowering	00	50	30	27	33	37	7	23	67
<i>Lamium amplexicaule</i>	Flowering	10	37	37	60	100	100	13	20	30
<i>Salvia spicata</i> L.	Flowering	10	70	30	67	93	97	10	17	67
Leguminosae										
<i>Cytisus scutellari</i> L.	Flowering	33	97	97	77	77	67	33	13	40
<i>Cytisus arvensis</i> L.	Vegetative	---	---	---	50	90	53	33	13	10
Umbelliferae										
<i>Galium aparine</i> L.	Flowering	---	---	---	30	60	63	10	20	47
<i>Matricaria inodora</i> (L.) W. Greuter	Vegetative	30	60	90	---	---	---	37	10	90
Compositae										
<i>Wasson discolor</i> Seb.	Pre-flowering	30	10	30	20	23	27	37	17	20
Malvaceae										
<i>Malva sylvestris</i> L.	Pre-flowering	30	60	90	---	---	---	33	37	77
Rapaceae										
<i>Helianthus annuus</i> L.	Flowering	33	50	50	57	100	100	33	10	77
Rubiaceae										
<i>Scorpiurus maritimus</i> L.	Flowering	00	50	77	30	53	30	37	10	77
<i>Scorpiurus maritimus</i> L.	Flowering	---	---	---	60	100	100	33	10	40
Pentagraceae										
<i>Plantago lanceolata</i> L.	Flowering	00	10	60	67	70	70	30	37	13
Polygonaceae										
<i>Polygonum aviculare</i> L.	Early flowering	20	90	93	---	---	---	---	---	---
<i>Rumex crispus</i>	Vegetative	07	60	83	67	63	30	37	37	17
Primulaceae										
<i>Anagallis arvensis</i> L.	Flowering	63	100	100	13	17	87	00	13	17
<i>Anagallis borealis</i> M.	Flowering	00	77	80	67	77	87	00	13	63
Ranunculaceae										
<i>Ranunculus acris</i> L.	Flowering	00	30	00	00	00	00	00	00	00
Rutaceae										
<i>Rutaceae aurantiacum</i> L.	Flowering	---	---	---	63	43	60	30	30	34
Rubiacae										
<i>Galium aparine</i> L.	Early flowering	37	50	70	---	---	---	20	20	33
Scrophulariaceae										
<i>Salvia officinalis</i> L.	Flowering	30	43	77	13	10	13	30	13	27
Umbelliferae										
<i>Anthriscus</i> L.	Flowering	37	70	80	67	100	100	33	37	70
<i>Conium maculatum</i>	Vegetative	30	70	87	30	100	100	37	13	77
<i>Thalictrum flavum</i> L.	Pre-flowering	77	87	90	57	77	87	13	20	77
<i>Heracleum sphacelatum</i> L.	Pre-flowering	00	100	100	60	70	77	17	67	87
<i>Heracleum sphacelatum</i> L.	Vegetative	37	80	90	77	80	90	30	10	50
<i>Saxifraga hypnoides</i> L.	Flowering	10	67	80	50	80	90	00	13	67
Urticaceae										
<i>Rumex crispus</i> L.	Early flowering	30	47	70	57	90	97	00	33	20
<i>Urtica dioica</i> L.	Flowering	17	77	87	97	90	93	33	37	40
USD (P > 0.5)										
		17	27	12	15	10	11	36	38	12

1-10) Score where the lowest score denotes no fungal growth while the highest score denotes that the Petri-dish was full of fungal growth

Table 1. Visual estimation of the effect of shoot extracts of different weed species on the growth of *P. digitatum*, *S. Sclerotiorum* and *V. dahliae* at three dates of incubation [Nomenclature is that of Flora Palaestina, Zohary (1966)] (continued)

Form and Scientific name of weeds	Growth stage	<i>P. digitatum</i>			<i>S. sclerotiorum</i>			<i>V. dahliae</i>		
		Days of incubation			Days of incubation			Days of incubation		
		4	8	16	4	8	16	4	8	16
Labiatae										
<i>Bellota saxatilis</i> C. Presl	Vegetative	1.7	9.0	9.3	8.0	9.3	9.3	0.0	1.3	1.7
<i>Lamium</i> sp.	Flowering	0.3	6.3	9.0	2.7	3.3	3.7	1.7	2.3	6.7
<i>Lamium amplexicaule</i> L.	Flowering	1.3	9.7	9.7	8.0	10.0	10.0	1.3	5.0	3.0
<i>Salvia syriaca</i> L.	Flowering	1.3	7.3	9.3	8.7	9.3	9.7	1.3	1.7	8.7
Leguminosae										
<i>Lupinus varius</i> L.	Flowering	3.3	9.7	9.7	7.7	7.7	9.7	0.3	1.3	4.0
<i>Cytisus arvensis</i> L.	Vegetative	--	--	--	3.0	9.0	9.3	0.0	1.3	1.3
Liliaceae										
<i>Bellevalia densiflora</i> Boiss	Flowering	--	--	--	6.0	8.0	8.3	1.0	2.0	8.7
<i>Muscana racemosa</i> (L.) Mill	Vegetative	0.0	8.0	9.0	---	---	---	0.7	1.0	5.3
Loasaliaceae										
<i>Nectandra cristatum</i> Sieb	Pre-flowering	0.0	1.3	3.0	2.3	2.3	2.7	0.7	1.7	2.0
Moraceae										
<i>Morva sp.</i>	Pre-flowering	3.0	8.0	9.0	---	---	---	0.3	0.7	7.7
Papaveraceae										
<i>Papaver rhoeas</i> L.	Flowering	2.3	5.0	5.3	5.7	10.0	10.0	0.3	1.0	7.7
Papilionaceae										
<i>Scorpisus mucicatus</i> L.	Flowering	0.0	5.0	7.7	9.0	9.3	9.3	0.7	1.0	7.7
<i>Vicia narbonensis</i> L.	Flowering	---	---	---	6.0	10.0	10.0	0.3	1.3	4.3
Plantaginaceae										
<i>Plantago lanceolata</i> L.	Flowering	0.0	1.3	6.0	8.7	7.0	7.0	0.3	0.7	1.3
Polygonaceae										
<i>Polygonum aviculare</i> L.	Early flowering	2.0	9.0	9.3	---	---	---	---	---	---
<i>Rumex crispus</i> L.	Vegetative	0.7	6.0	8.3	6.7	8.3	9.0	0.7	0.7	1.7
Primulaceae										
<i>Anagallis arvensis</i> L.	Flowering	8.3	10.0	10.0	7.3	7.7	8.7	0.0	1.3	1.7
<i>Anagallis foemina</i> Mill	Flowering	0.0	7.7	8.0	6.7	7.7	8.7	0.0	1.3	6.3
Ranunculaceae										
<i>Ranunculus asiaticus</i> L.	Flowering	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Rosaceae										
<i>Rosula sp.</i>	Flowering	--	--	---	6.0	8.3	9.0	0.0	0.3	0.4
Rubiaceae										
<i>Galium aparine</i> L.	Early flowering	0.7	5.0	7.0	---	---	---	2.0	2.3	3.3
Solanaceae										
<i>Solanum elaeagnifolium</i> L.	Fruiting	0.0	4.3	7.7	1.3	1.3	1.3	0.0	1.3	2.7
Umbelliferae										
<i>Anthriscus</i> L.	Flowering	0.7	7.3	8.3	8.7	10.0	10.0	0.3	0.7	7.3
<i>Conium maculatum</i>	Vegetative	0.0	7.3	8.7	9.0	10.0	10.0	0.7	1.3	7.7
<i>Falcaria vulgaris</i> Bernh	Pre-flowering	7.7	8.7	9.3	5.7	7.7	8.7	1.3	2.0	7.7
<i>Ferula communis</i> L.	Pre-flowering	9.0	10.0	10.0	6.3	7.0	7.7	1.7	8.7	8.7
<i>Foeniculum vulgare</i> L.	Vegetative	0.7	8.3	9.0	7.7	8.3	9.3	0.0	1.0	5.3
<i>Sonchus oleraceus</i> L.	Flowering	1.0	6.7	8.3	5.0	8.0	9.0	0.0	1.3	6.7
Urticaceae										
<i>Urtica dioica</i> (L.) Koch	Early flowering	0.0	4.7	7.3	5.7	9.3	9.7	0.0	0.3	2.3
<i>Urtica urens</i> L.	Flowering	1.7	7.7	8.7	8.7	9.3	9.3	0.3	0.7	4.3
LSD (P<0.5)		1.7	2.1	1.2	1.5	1.3	1.1	0.8	0.8	1.2

1-10: Scale where the lowest score denotes no fungal growth while the highest score denotes that the Petri-dish was full of fungal growth

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Family and Scientific name of weeds	Growth stage	<i>P. digitatum</i>			<i>S. sclerotiorum</i>			<i>V. dahliae</i>		
		Days of incubation			Days of incubation			Days of incubation		
		4	8	16	4	8	16	4	8	16
Labiatae										
<i>Ballota saxatilis</i> C. Presl	Vegetative	1.7	9.0	9.3	8.0	8.3	9.3	0.0	1.3	1.7
<i>Lamium</i> sp.	Flowering	0.3	6.3	8.0	2.7	3.3	3.7	1.7	2.3	6.7
<i>Lamium amplexicaula</i> L.	Flowering	1.3	9.7	9.7	8.0	10.0	10.0	1.3	3.0	3.0
<i>Salvia syriaca</i> L.	Flowering	1.3	7.3	8.3	8.7	9.3	9.7	1.3	1.7	8.7
Leguminosae										
<i>Lupinus varius</i> L.	Flowering	3.3	9.7	9.7	7.7	7.7	9.7	0.3	1.3	4.0
<i>Ononis antiquorum</i>	Vegetative	---	---	---	3.0	9.0	9.3	0.0	1.3	1.3
Liliaceae										
<i>Bellevalia densiflora</i> Boiss	Flowering	---	---	---	8.0	8.0	8.3	1.0	2.0	8.7
<i>Muscaria racemosum</i> (L.) Mill	Vegetative	0.0	8.0	9.0	---	---	---	0.7	1.0	5.3
Loranthaceae										
<i>Viscum cruciatum</i> Sieb	Pre-flowering	0.0	1.3	3.0	2.3	2.3	2.7	0.7	1.7	2.0
Malvaceae										
<i>Malva sylvestris</i> L.	Pre-flowering	3.0	8.0	9.0	---	---	---	0.3	0.7	7.7
Papaveraceae										
<i>Papaver rhoeas</i> L.	Flowering	2.3	5.0	5.3	5.7	10.0	10.0	0.3	1.0	7.7
Papilionaceae										
<i>Scorpiurus muricatus</i> L.	Flowering	0.0	5.0	7.7	9.0	9.3	9.3	0.7	1.0	7.7
<i>Vivicia narbonensis</i> L.	Flowering	---	---	---	6.0	10.0	10.0	0.3	1.3	4.3
Plantaginaceae										
<i>Plantago lanceolata</i> L.	Flowering	0.0	1.3	6.0	6.7	7.0	7.0	0.0	0.7	1.3
Polyginaceae										
<i>Polygonum aviculare</i> L.	Early flowering	2.0	9.0	9.3	---	---	---	---	---	---
<i>Rumex crispus</i> L.	Vegetative	0.7	6.0	8.3	6.7	8.3	9.0	0.7	0.7	1.7
Primulaceae										
<i>Anagallis arvensis</i> L.	Flowering	8.3	10.0	10.0	7.3	7.7	8.7	0.0	1.3	1.7
<i>Anagallis foemina</i> Mill	Flowering	0.0	7.7	8.0	6.7	7.7	8.7	0.0	1.3	6.3
Ranunculaceae										
<i>Ranunculus asiaticus</i> L.	Flowering	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Rosaceae										
<i>Poterium spinosum</i> L.	Flowering	---	---	---	6.3	8.3	9.0	0.0	0.3	0.4
Rubiaceae										
<i>Galium tricoratum</i> Dandy	Early flowering	0.7	5.0	7.0	---	---	---	2.0	2.3	3.3
Solanaceae										
<i>Solanum nigrum</i> L.	Fruiting	0.0	4.3	7.7	1.3	1.3	1.3	0.0	1.3	2.7
Umbelliferae										
<i>Ammi majus</i> L.	Flowering	0.7	7.3	8.3	8.7	10.0	10.0	0.3	0.7	7.3
<i>Conium maculatum</i>	Vegetative	0.0	7.3	8.7	9.0	10.0	10.0	0.7	1.3	7.7
<i>Falcaria vulgaris</i> Bernh	Pre-flowering	7.7	8.7	9.3	5.7	7.7	8.7	1.3	2.0	7.7
<i>Ferula communis</i> L.	Pre-flowering	9.0	10.0	10.0	6.3	7.0	7.7	1.7	8.7	8.7
<i>Foeniculum vulgare</i> L.	Vegetative	0.7	8.3	9.0	7.7	8.3	9.3	0.0	1.0	5.3
<i>Sandix pecten-veneris</i> L.	Flowering	1.0	6.7	8.3	5.0	8.0	9.0	0.0	1.3	6.7
Urticaceae										
<i>Parietaria diffusa</i> (L.) Koch	Early flowering	0.0	4.7	7.3	5.7	9.3	9.7	0.0	0.3	2.3
<i>Urtica urens</i> L.	Flowering	1.7	7.7	8.7	8.7	9.3	9.3	0.3	0.7	4.3
LSD (P<0.5)		1.7	2.1	1.2	1.5	1.3	1.1	0.8	0.8	1.2

1-10 : Scale where the lowest score denotes no fungal growth while the highest score denotes that the Petri-dish was full of fungal growth

Out of 62 species tested for fungitoxic effects against this fungus, extracts of more than 40 weed species significantly reduced its colony growth compared with control. However, extracts of *A. arvensis*, *A. leucoclada*, *B. saxatilis*, *B. didyma*, *G. tricornutum*, *I. viscosa*, *L. amplexicaula*, *O. antiquorum*, *P. diffusa*, *P. spinosum*, *R. asiaticus*, *S. nigrum* and *V. cruciatum* were the most phytotoxic to the fungus and caused more than 60% growth inhibition compared with the control.

Fungitoxic effect of extracts on spore formation of *P. digitatum* and *V. dahliae* is shown in Table 2. Differences in the effect of various extracts on sporulation of *P. digitatum* were not clear at 4 days of incubation. However, at 8 days, they were 18 species inhibitory to spore formation and 24 species at 16 days of incubation. *C. murale*, *C. aspera*, *P. lanceolata*, *R. asiaticus*, *S. irio* and *V. cruciatum* extracts severely reduced sporulation of this fungus compared with the control.

The effect of different extracts on sporulation of *V. dahliae* was clear at 4 days of incubation at which extracts of 40 weed species inhibited its spore formation. At 16 days of incubation period, there were 46 inhibitory weed species. The most effective extracts were those of *A. arvensis*, *B. saxatilis*, *O. antiquorum*, *P. lanceolata* and *R. asiaticus*.

DISCUSSION

The study of the fungitoxic effects of aqueous shoot extracts from different weed species indicate the importance of many weed species as a possible natural source of fungitoxic materials (Tables 1 & 2).

Certain weed extracts inhibited growth and /or sporulation of one more fungi species tested. Antifungal effects of extracts of many plant species have been reported (Frange, 1984 ; Al Abed *et al.*, 1993 ; Singh, 1994). In the present study, extracts of some of the most common weed species showed antifungal activity. Among those were *A. arvensis*, *C. murale*, *I. viscosa*, *S. nigrum* and *R. asiaticus*.

Fungitoxicity of extracts of certain *Chenopodium* species against other plant pathogens have been reported (Dubey *et al.*, 1983 ; Rafik *et al.*, 1984), while results obtained with *R. asiaticus* confirmed the importance of *Ranunculus* species as plants

exhibiting antifungal properties as *R. sclertatus*, *R. bulbosus* and *R. clematis* have also been reported to have antifungal properties (Misra *et al.*, 1974 ; Misra & Dixit, 1978 ; Mares, 1987).

Antifungal activity of *A. arvensis* and *I. viscosa* extracts against *Helminthosporium sativum* and *Fusarium oxysporum* have been also reported (Al Abed *et al.*, 1993). Results showed that extracts of different weed species varied in their antifungal potential. Such differences were to be expected since plants varied in their chemical constituent, habitats and growth stages at which they were collected. Differences in the nature and/or concentration of inhibitory materials even between different plant parts were found (Frange, 1984 ; Al Abed (1992).

Environmental conditions and growth habitats may greatly influence the production and concentration of inhibitory materials and the ability of plants to release them under stress conditions (Rice, 1984). Differences were also noticed among species of the same family as well as those of different families.

This may be due to differences in the nature and concentration of inhibitory chemicals between different plant species or to differences in their ability to diffuse through the growing medium and subsequently to inhibit fungus growth. Other researchers reported differences in the antifungal activities from species to species, genus to genus and from family to another (Al Abed *et al.*, 1993 ; Rizki *et al.*, 1984).

Extracts of certain weed species enhanced growth and/or sporulation of the fungus compared with the control. This stimulatory effect may be due to the presence of growth promoting substances or nutrients in their extracts or to the stimulatory effect of growth inhibitors found at low concentration in plant tissues. Other researchers reported stimulatory effect of different plant extracts especially when low concentrations were used (Rizki *et al.*, 1984 ; Shakhawat & Prasada, 1971).

The antifungal effect of extracts reduced with incubation period for all species screened except that of *R. asiaticus*. This fate of chemical inhibitors may be due to the transformation of these materials to non toxic forms or the loss of some volatile inhibitors during the relatively long incubation period. Volatile materials of different plant species have been reported to have antifungal properties

Table 2. Visual estimation of the effect of shoot extracts of different weed species on the sporulation of *P. digitatum* and *V. dahliae* et three dates of incubation

Weed species	<i>P. digitatum</i>			<i>V. dahliae</i>		
	Days of incubation			Days of incubation		
	4	8	16	4	8	16
H ₂ O (control)	0.3	3.3	7.7	3.0	4.0	9.7
Aristolochiaceae						
<i>Aristolochia maurorum</i>	0.0	4.0	7.0	1.0	1.7	5.7
Asclepiadaceae						
<i>Calotropis procera</i>	0.3	6.0	6.7	1.0	3.0	8.7
Boraginaceae						
<i>Symphytum palaestinum</i>	0.0	6.3	7.3	0.0	0.3	4.3
Caryophyllaceae						
<i>Cerastium dicotomum</i>	0.0	5.7	7.7	0.7	1.7	7.0
<i>Stellaria media</i>	5.7	6.7	8.7	2.3	2.3	8.3
Chenopodiaceae						
<i>Atriplex leucoclada</i>	0.0	1.0	8.0	0.0	1.0	4.0
<i>Chenopodium murale</i>	0.6	0.3	2.3	2.0	2.7	9.3
Compositae						
<i>Anthemis cotula</i>	---	---	---	2.0	2.7	9.3
<i>Calendula arvensis</i>	0.0	5.3	7.7	2.3	3.3	9.3
<i>Carthamus nitidus</i>	0.6	3.7	6.7	2.7	3.7	4.7
<i>Centaurea iberica</i>	---	---	---	1.7	4.0	9.3
<i>Crepis aspera</i>	0.0	0.3	2.3	2.0	3.0	8.0
<i>Gendelia tournefortii</i>	0.0	1.7	6.0	2.0	4.3	8.3
<i>Inula viscosa</i>	0.0	0.6	6.3	1.3	2.7	3.7
<i>Notobasis syriaca</i>	0.0	1.7	4.7	1.3	3.0	10.0
<i>Onopordum jordanicolum</i>	0.0	4.7	6.7	0.3	1.3	7.7
<i>Senecio vernalis</i>	0.0	1.7	8.7	0.3	1.0	7.7
<i>Sonchus oleraceus</i>	0.0	2.0	6.0	1.0	3.3	5.3
Convolvulaceae						
<i>Convolvulus althaeoides</i>	0.0	3.7	6.7	0.7	1.7	7.0
<i>Convolvulus arvensis</i>	0.3	4.7	7.3	2.7	3.0	7.7
Cruciferae						
<i>Biscuttela didyma</i>	0.0	2.7	7.3	2.7	3.3	3.7
<i>Capsella bursa-pastoris</i>	0.0	2.7	6.0	0.3	1.3	6.3
<i>Cardaria draba</i>	0.0	2.0	5.0	1.3	3.0	9.3
<i>Diplotaxis erucoides</i>	0.3	7.0	7.7	3.0	3.7	9.0
<i>Eruca sativa</i>	0.3	4.0	6.3	2.0	3.7	5.3
<i>Erucaria hispanica</i>	0.0	1.3	6.0	0.7	1.7	7.3
<i>Sinapis arvensis</i>	0.7	4.3	6.3	1.7	2.0	8.7
<i>Sisymbrium irio</i>	0.0	1.3	2.7	1.0	2.7	8.3
Cucurbitaceae						
<i>Ecballium elaterium</i>	0.0	1.0	6.3	2.0	3.0	8.3
Euphorbitaceae						
<i>Euphorbia helioscopia</i>	2.0	4.3	7.3	1.3	1.7	5.0
<i>Mercurialis annua</i>	0.0	1.0	5.7	2.0	3.0	9.7
Fumariaceae						
<i>Fumaria densiflora</i>	0.0	1.0	5.0	2.7	3.3	6.3
Geraniaceae						
<i>Erodium cruciatum</i>	---	---	---	1.0	2.3	8.3
Graminae						
<i>Avena sterilis</i>	0.0	1.0	7.3	---	---	---

Table 2. Visual estimation of the effect of shoot extracts of different weed species on the sporulation of *P. digitatum* and *V. dahliae* at three dates of incubation (Continued)

Weedspecies	<i>P. digitatum</i>			<i>V. dahliae</i>		
	Days of incubation			Days of incubation		
	4	8	16	4	8	16
Labiatae						
<i>Ballota saxatilis</i>	0.3	7.0	7.3	0.7	2.3	3.0
<i>Lamium</i> sp.	0.0	1.0	6.7	3.3	4.3	7.3
<i>Lamium amplexicaula</i>	0.3	7.0	8.3	1.3	2.3	4.7
<i>Salvia syriaca</i>	0.3	1.7	7.0	2.7	3.0	9.7
Leguminosae						
<i>Lupinus varius</i>	0.0	5.3	7.7	2.0	2.7	5.3
<i>Oninis antiquorum</i>	---	---	---	0.0	0.7	2.7
Liliaceae						
<i>Bellevalia densiflora</i>	---	---	---	3.3	3.7	10.0
<i>Muscaria racemosum</i>	0.0	0.6	7.0	0.7	1.3	5.7
Loranthaceae						
<i>Viscum cruciatum</i>	0.0	0.3	2.3	2.3	3.0	4.0
Malvaceae						
<i>Malva sylvestris</i>	0.0	4.7	7.7	0.7	1.7	9.7
Papaviraceae						
<i>Papaver rhoeas</i>	0.3	4.7	4.7	1.7	4.0	8.3
Papilionaceae						
<i>Scorpiurus muricatus</i>	0.0	1.0	4.3	1.3	2.0	9.3
<i>Vicia narbonensis</i>	---	---	---	1.0	2.0	5.3
Plantaginaceae						
<i>Plantago lanceolata</i>	0.0	0.3	3.3	0.0	0.7	3.3
Polygonaceae						
<i>Polygonum aviculare</i>	0.0	2.7	8.0	---	---	---
<i>Rumex crispus</i>	0.0	3.0	4.3	1.7	2.7	8.0
Primulaceae						
<i>Anagallis arvensis</i>	6.0	6.7	8.7	0.0	0.7	3.3
<i>Anagallis foemina</i>	0.0	4.0	6.7	0.7	1.0	7.7
Ranunculaceae						
<i>Ranunculus asiaticus</i>	0.0	0.0	0.0	0.0	0.0	0.0
Rosaceae						
<i>Poterium spinosum</i>	---	---	---	0.7	3.0	4.7
Rubiaceae						
<i>Galium tricornutum</i>	0.3	7.7	8.3	2.0	3.0	4.3
Solanaceae						
<i>Solanum nigrum</i>	0.0	6.7	6.7	0.7	2.0	4.0
Umbelliferae						
<i>Ammi majus</i>	0.0	1.0	6.7	1.3	2.0	8.0
<i>Conium maculatum</i>	0.3	2.3	4.7	1.3	3.0	8.7
<i>Falcaria vulgaris</i>	8.3	8.3	8.3	2.3	2.3	8.0
<i>Ferula communis</i>	0.3	6.7	7.7	2.3	3.3	9.7
<i>Foeniculum vulgare</i>	0.0	6.7	7.3	1.7	3.3	6.3
<i>Scandix pecten- veneris</i>	0.6	1.0	6.0	0.7	2.3	7.0
Urticaceae						
<i>Parietaria diffusa</i>	0.0	2.3	5.3	1.7	3.0	3.7
<i>Urtica uren</i>	0.0	5.0	7.3	1.3	3.0	5.3
LSD (P <0.5)	1.1	1.8	1.2	1.2	1.1	1.2

1- 10 Scale where the lowest score denotes no fungal spores while the highest denotes that the petridish was full of fungal spores.

(Al Abed, 1992) and reduction in the phytotoxicity of plant extract with time has been well documented (Al Abed *et al.*, 1993 ; Dumancas, 1976 ; Chaturvedi *et al.*, 1987).

However, the persistence of inhibitory effect of *R. asiaticus* and to a less extent those of *C. murale* and *C. aspera* may be due to the high stability of inhibitory chemicals and less effect of incubation temperature on the activity of these inhibitors.

The effect of extracts on spore formation of the fungus is a useful indicator on the toxic effect they have against these fungus. This effect is of great importance for inhibiting fungi development and preventing them from successfully completing their life cycles.

Extracts of certain species appeared to be highly selective against certain fungus while others such as *R. asiaticus* showed general effects. This confirm the previous speculation that the nature of inhibitory materials is different between species and reflects differences in their antifungal effects.

Plants were extracted in water and thus it is more likely that only water soluble inhibitors were dissolved. The solubility of chemical inhibitors of different weed species in water may be the reason behind the differences obtained in the antifungal activity of different extracts.

However, there is a good indication that many weed species contain water soluble antifungal material(s), and these could be easily extracted in water. This, however, did not negate the possible presence of other chemical inhibitors in plant tissues not soluble in water.

Our study was a preliminary one and as such avoided investigation using different organic solvents with their consequent complications.

CONCLUSION

This study showed that many weed species contain water soluble chemicals of antifungal properties in their tissues. Some of these proved to be good inhibitors to growth and/or sporulation one or more fungi species.

Extracts of *R. asiaticus* appeared to be highly effective against the three fungi and gave almost complete growth and sporulation inhibition. There is a great potential remained to consider this study

as a base to substantiate further work on the promising extracts for further understanding the role that plants may play in the future for developing more safe and new natural pesticides.

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REFERENCES

- Akhtar T., Sattar A. & Khan I. (1986) Antifungal activity of some plant extracts against potato dry rot of *Fusarium*. *Sarbad Journal of Agriculture* 2:187-191
- Al Abed A.S. (1992) Possible antifungal effects of aqueous extracts and residues of some common wild plant species on certain plant pathogenic fungi. M. Sc. Thesis, University of Jordan, pp. 81
- Al Abed A.S., Qasem J.R. & Abu-Blan H.A. (1993) Antifungal effects of some common wild plant species on certain plant pathogenic fungi. *Dirasat (Pure and Applied Science)* 20 : 149-158
- Beye F. (1978) Insecticides from the vegetable Kingdom. *Plant Research and development* 7 : 13-31
- Chaturvedi R., Dikshit A. & Dixit S.N. (1987) *Adenocalymma allicea*. A new source of a natural fungitoxicant. *Tropical Agriculture* 64 : 318-322
- Dubey N.K., Kishore N., Srivastava O.P., Dikshit A. & Singh S.K. (1983) Fungitoxicity of some higher plants against *Rhizoctonia solani*. *Plant and Soil* 72 : 91-94
- Dumancas E.E. (1976) A survey of the higher plants for fungicidal properties against *Helminthosporium oryzae* Breda de Haan and *Pyricularia oryzae* Cav. M. Sc. Thesis, University of Philippines at Los Bafios, College, Laguna. 93p.
- Frangé N.S. (1984) Evaluation of medicinal plant extracts as protectant and therapeutant against legume pathogens. Ph.D Thesis, University of Philippines
- Guesin J.C. & Reveillere H. (1984) Antifungal activity of plant extract against 9 fungi species. *Annuaire Pharmaceutique Français* 42 : 553-559
- Lapis D.B. & Dumanca E.E. (1978) Fungicidal activity of crude extracts against *Helminthosporium oryzae*. *Philippines Phytopathology* 14 : 23-37

- Macias F. A., Vrela R.M., Torres A. & J. M.G. Mollinillo (1994) Potential of sunflower (*Helianthus annuus* L.) as source of natural herbicides models. *International Symposium, Allelopathy in Sustainable Agriculture, Forestry and Environment*. New Delhi, Sept. 6-8 P68 (Abstract)
- Mares D. (1987) Antimicrobial activity of protoanemonin, a lacton from ranunculaceous plants. *Mycopathologia* 98 (3) : 133-14
- Misra S. B. & Dixit S.N. (1978) Antifungal properties of leaf extract of *Ranunculus scleratus* L. *Experientia* 34 : 1442-1443
- Misra S.B., Misra R.B.R. & Dixit S.N. (1974) Screening of higher plants for antifungal activity. *National Academy of Science (India)* 76 : 203-205
- Narwal S.S. (1994) Allelopathy in crop production, Scientific Publishers, Jodhpur, India, pp. 288
- Rafiq M., Nasir M.A. & Bhatti M.A.R. (1984) antifungal properties of certain common wild plants against different fungi. *Pakistan Journal of Agriculture Research* 5 : 236-238
- Rice E.L. (1984) Allelopathy. 2nd Eddition. Acedemic Press London.
- Rice E.L. (1986) Pest Control with Nature's chemicals. University of Oklahoma Press. Norman, USA
- Rizki Y.M., Fatima K., Ahmed I. & Yasmin B. (1984) Studies on antifungal properties of indigenous plants from Karachi region. Part II. *Pakistan Journal of Science Research* 32 : 608-611.
- Shekhawat P.S. & Prasada R. (1971) Antifungal properties of some plant extracts. I. Inhibition of spore germination. *Indian Phytopathology* 24 : 800-802
- Singh D. (1994) Scope of medecinal and aromatic plants in pest management. *International Symposium, Allelopathy in Sustainable Agriculture, Forestry and Enviornment*. New Delhi, Sept. 6-8 P68 (Abstract)
- Zohary M. (1966) Flora Palaestina. The Israel Academy of Science and Humanities, Jerusalem.